

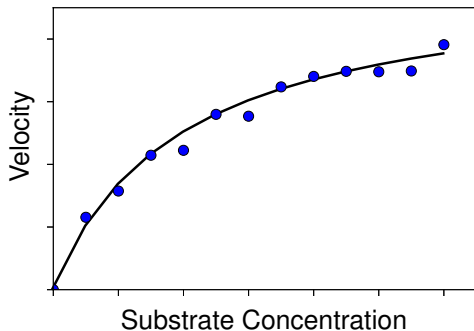
Biological Chemistry Laboratory  
Biology 3515/Chemistry 3515  
Spring 2018

Lecture 13:

Data Analysis for the  $V$  versus  $[S]$  Experiment  
and Interpretation of the Michaelis-Menten Parameters

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## Analysis of Data from the $V$ versus $[S]$ Experiment



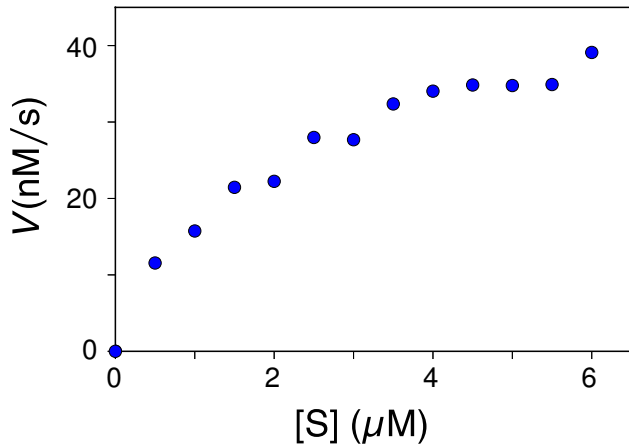
- We want to fit the experimental data to the Michaelis-Menten Equation:

$$V = \frac{[S]V_{\max}}{[S] + K_m}$$

- From the fit, we obtain estimates of  $K_m$  and  $V_{\max}$ .

# Clicker Question #1

Estimate  $V_{\max}$  from the graph:



1 30 nM/s

2 40 nM/s\*

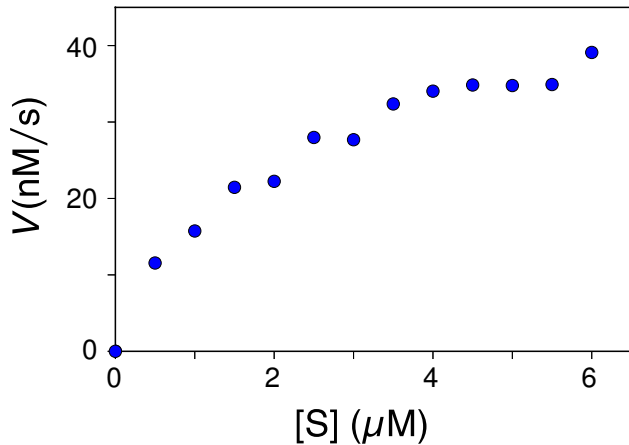
3 50 nM/s

4 80 nM/s

\*close enough for credit.

## Clicker Question #2

Estimate  $K_m$  from the graph:



1  $1 \mu\text{M}$

2  $2 \mu\text{M}$

3  $5 \mu\text{M}$

4  $10 \mu\text{M}$

# A Classic Method for Analyzing Enzyme Kinetics Data

- Rearrangement of the Michaelis-Menten Equation:

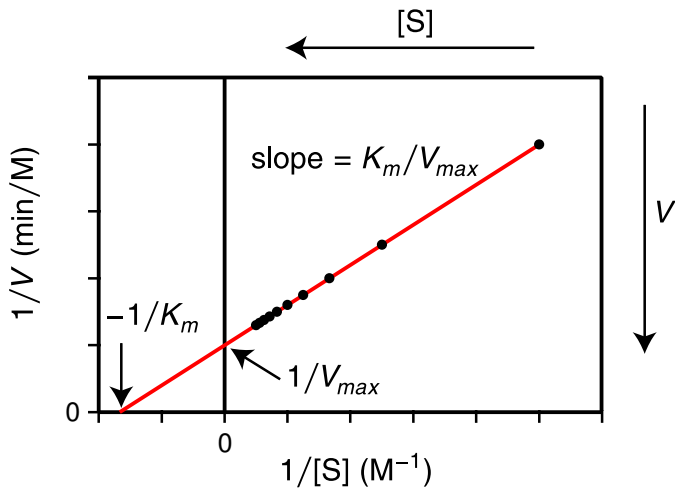
$$V = \frac{[S]V_{\max}}{[S] + K_m}$$

$$\frac{1}{V} = \frac{[S] + K_m}{[S]V_{\max}} = \frac{[S]}{[S]V_{\max}} + \frac{K_m}{[S]V_{\max}}$$

$$\frac{1}{V} = \frac{1}{[S]} \cdot \frac{K_m}{V_{\max}} + \frac{1}{V_{\max}}$$

- A plot of  $1/V$  versus  $1/[S]$  should generate a straight line with a slope of  $K_m/V_{\max}$  and an intercept of  $1/V_{\max}$  on the  $1/V$  axis.

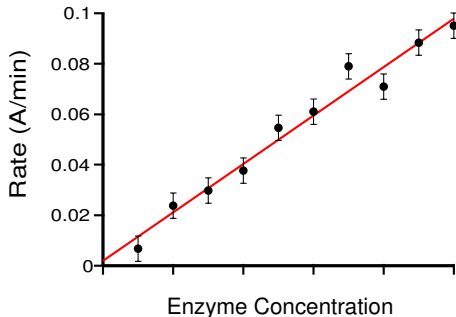
# The Lineweaver-Burk Plot



- If the data are perfect, this plot gives good estimates of  $K_m$  and  $V_{max}$ .
- But, experimental error in  $V$  can lead to strange effects!

# Experimental Error and Uncertainty

- Error bars for rate measurements are of approximately constant size (e.g.,  $\pm 0.005$  A/min), rather than a constant percentage of the measurement.



- For 0.1 A/min,  $\pm 0.005$  A/min =  $\pm 5\%$ .
- For 0.01 A/min,  $\pm 0.005$  A/min =  $\pm 50\%$ .
- Least-squares fitting works well if the *absolute* uncertainties of all data points are approximately equal.

# What Happens When We Take Reciprocals?

- $V = 0.1 \pm 0.005$

$$\frac{1}{0.105} = 9.52,$$

$$\frac{1}{0.095} = 10.5,$$

$$\frac{1}{V} = 10 \pm 0.5$$

- $V = 0.01 \pm 0.005$

$$\frac{1}{0.015} = 66.7,$$

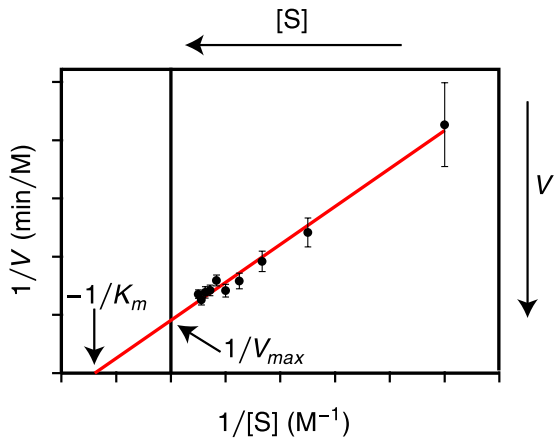
$$\frac{1}{0.005} = 200,$$

$$\frac{1}{V} = 100 \pm 50$$

- The values of  $1/V$  derived from small velocities can have huge absolute errors.



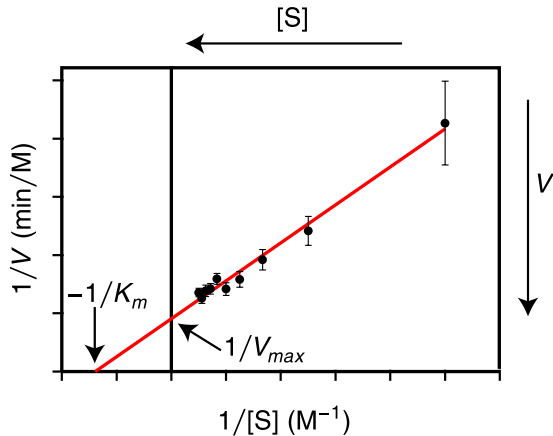
## The Effects on a Lineweaver-Burk Plot



- Errors in the least precise measurements (low  $V$ ) can cause large changes in the line fit to the Lineweaver-Burk plot.

## Clicker Question #3

Which parameter is likely to be more sensitive to errors in a Lineweaver-Burk plot?



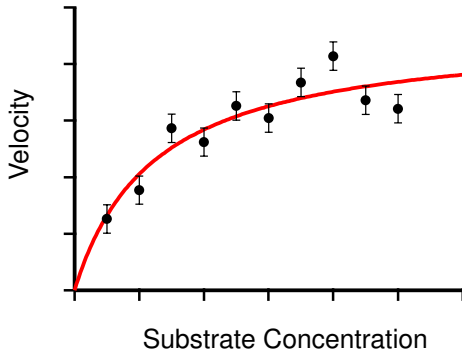
1  $K_m$ \*

2  $V_{max}$

\* Provided that values of  $V$  approach  $V_{max}$ .

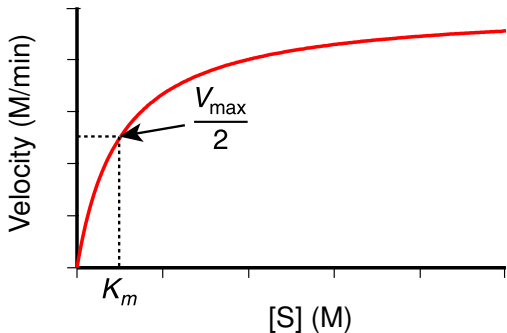
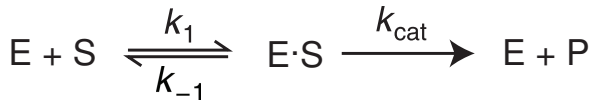
## Two Ways to Deal with This Problem

- Use Lineweaver-Burk, but weight data according to uncertainties in  $1/V$ .
- Fit velocity data directly to the Michaelis-Menten equation using non-linear least-squares method.



- Equal errors in  $V$  are weighted equally.

## Interpreting $K_m$



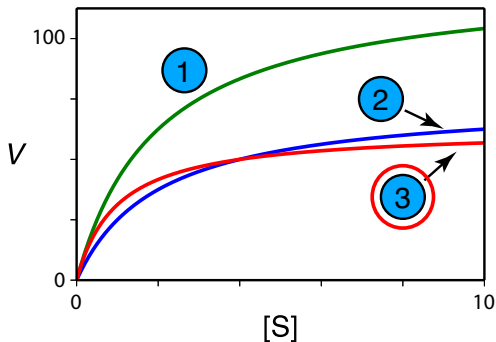
$$K_m = \frac{k_{-1} + k_{\text{cat}}}{k_1}$$

$$V = \frac{V_{\text{max}}}{2} \quad \text{when} \quad [S] = K_m$$

- When  $[S] = K_m$ , half of total enzyme has substrate bound.
- The larger  $K_m$  is, the more substrate is required to reach  $V_{\text{max}}/2$ , or any specified fraction of  $V_{\text{max}}$ .

## Clicker Question #4:

Data for three substrates with the same enzyme.



Which substrate binds most tightly to the enzyme?

No wrong answers, for now.

## A Closer Look at Binding and $K_m$ : $K_m$ versus $K_d$



- $K_m$  is defined in terms of the rate constants:

$$K_m = \frac{k_{-1} + k_{\text{cat}}}{k_1}$$

- $K_d$  is the equilibrium constant for dissociation.

$$K_d = \frac{[E][S]}{[E \cdot S]} = \frac{k_{-1}}{k_1}$$

A large  $K_d$  indicates weak binding.

## $K_m$ versus $K_d$

$$K_m = \frac{k_{-1} + k_{\text{cat}}}{k_1}$$

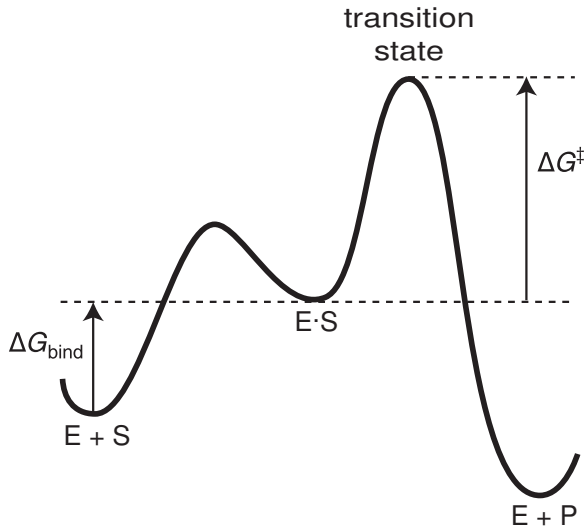
$$K_d = \frac{[E][S]}{[E \cdot S]} = \frac{k_{-1}}{k_1}$$

- If  $k_{\text{cat}} \ll k_{-1}$ , *i.e.*, the E·S complex is more likely to dissociate than undergo catalysis:

$$K_m \approx \frac{k_{-1}}{k_1} = K_d$$

- In general,  $K_m \geq K_d$
- Strength of equilibrium binding may be greater than indicated by  $K_m$ .

# Energy Profile for an Enzyme-Catalyzed Reaction



- Free-energy change for binding:

$$\Delta G_{\text{bind}} = RT \ln K_d$$

$R$  = Gas constant

$T$  = Temperature

- Free-energy change from E·S complex to transition state:

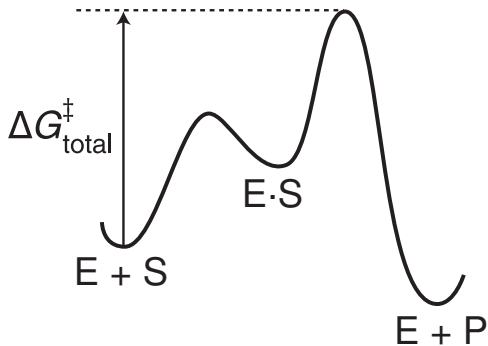
$$\Delta G^\ddagger = RT \ln \left( \frac{k_b T}{k_{\text{cat}} h} \right)$$

$k_b$  = Boltzmann constant

$h$  = Planck constant



# The Significance of $k_{\text{cat}}/K_m$



- Free-energy difference between  $E + S$  and the transition state:

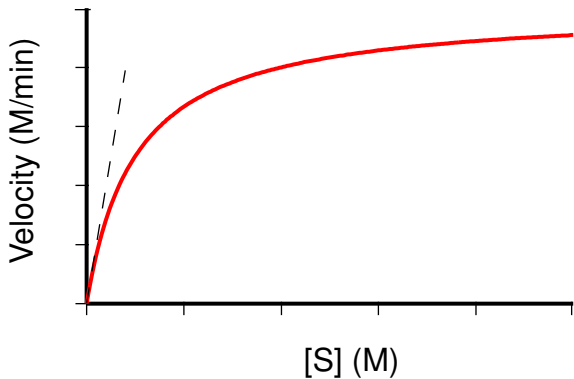
$$\Delta G_{total}^{\ddagger} = \underbrace{RT \ln \left( \frac{k_b T}{h} \right)}_{\text{Constant}} + RT \ln \left( \frac{K_d}{k_{\text{cat}}} \right)$$

- If  $k_{-1} \gg k_{\text{cat}}$ ,  $K_d \approx K_m$ :

$$\Delta G_{total}^{\ddagger} = C - RT \ln \left( \frac{k_{\text{cat}}}{K_m} \right)$$

- The ratio  $k_{\text{cat}}/K_m$  reflects the free energy difference between  $E + S$  and the transition state. (Assuming  $K_d \approx K_m$ )
- $k_{\text{cat}}/K_m$  is commonly interpreted as a measure of enzymatic efficiency.
- Catalytic efficiency is favored by a large value of  $k_{\text{cat}}$  and a small value of  $K_m$ .

# $k_{\text{cat}}/K_m$ Is Also the Apparent Second-Order Rate Constant at Low Substrate Concentrations



- If  $[S] \ll K_m$ :

$$V = \frac{[S][E]_T k_{\text{cat}}}{K_m + [S]}$$

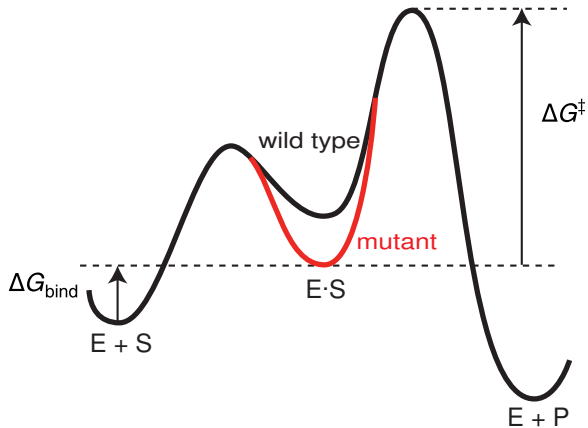
$$V \approx \frac{k_{\text{cat}}}{K_m} [S][E]_T$$

- Units of  $k_{\text{cat}}/K_m$

$$\frac{\text{s}^{-1}}{\text{M}} = \text{sec}^{-1}\text{M}^{-1}$$

## Is a Low $K_m$ Always Good?

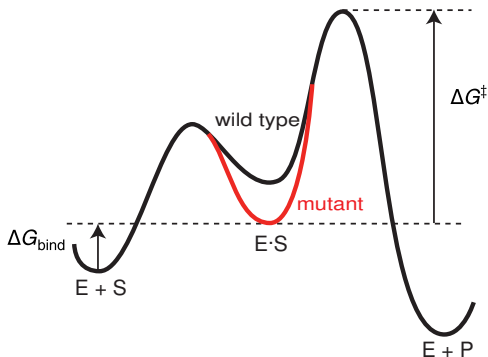
Suppose that we could design a mutant enzyme that forms a more stable complex with substrate.



This will lower  $K_m$  and  $k_{\text{cat}}$ , but leave  $k_{\text{cat}}/K_m$  the same.

## Clicker Question #5

At **low** substrate concentration ( $[S] \ll K_m$ ), will the velocity for the mutant enzyme be greater or less than that of the original enzyme?



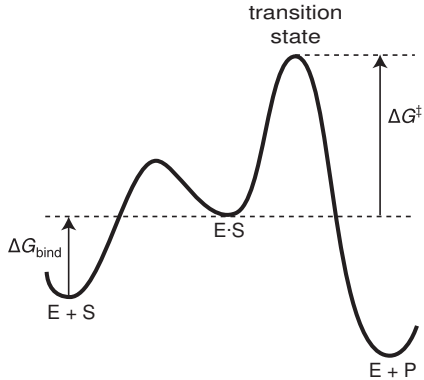
- 1 Greater than the original enzyme
- 2 Less than the original enzyme
- 3 The same as the original enzyme!

$$V = \frac{[S][E]_T k_{\text{cat}}}{K_m + [S]}$$

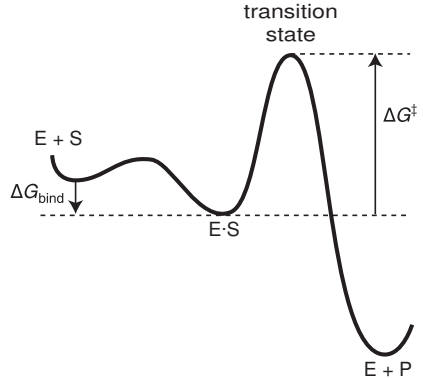
$$V \approx \frac{k_{\text{cat}}}{K_m} [S][E]_T$$

# What About High Substrate Concentrations?

Low Substrate Concentration



High Substrate Concentration



- When  $[S] \gg K_m$ , the enzyme-substrate complex is favored with respect to the free enzyme.