

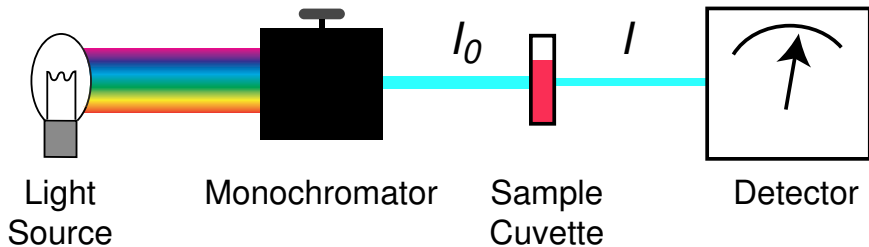
Biological Chemistry Laboratory
Biology 3515/Chemistry 3515
Spring 2017

Lecture 5

More on UV-visible Spectrophotometry:
Beer's Law and Overlapping Spectra

24 January 2017
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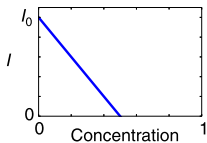
A UV-Visible Spectrophotometer



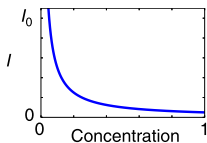
Clicker Question #1

How does transmitted light intensity change with concentration?

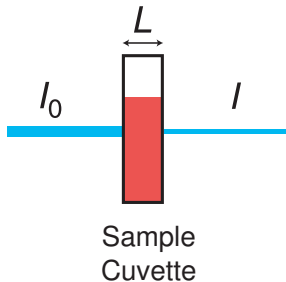
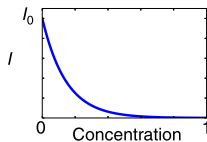
1 $I = I_0 - kC$



2 $I = \frac{I_0}{kC}$



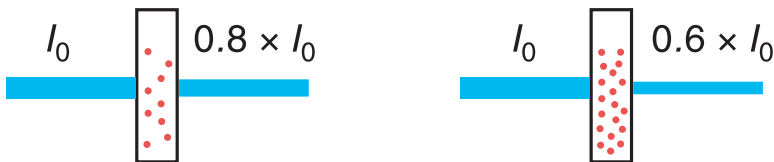
3 $I = I_0 e^{-kC}$



No wrong answers (for points)!

Clicker Question #2

If we double the concentration, does the number of photons absorbed double?



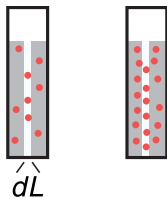
Click:

1 for Yes

2 for **No**

No wrong answers (for points)!

For a thin slice of the cuvette:



- The probability of a photon being absorbed is proportional to the concentration, C , and the thickness of the slice, dL :

$$p = kCdL$$

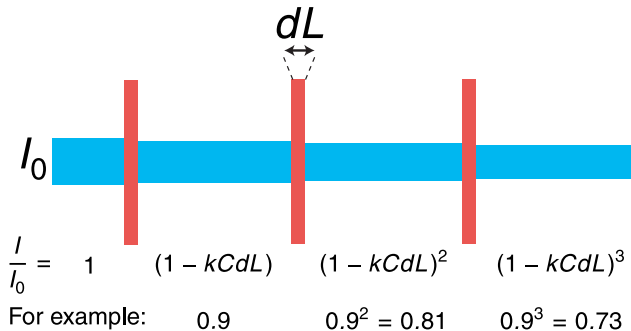
where k is a constant of proportionality.

- Fraction of light transmitted by a thin slice:

$$\frac{I}{I_0} = (1 - p) = (1 - kCdL)$$

- How thin does the slice have to be for this to be true?

Many Thin Slices



- For N slices:
$$\frac{I}{I_0} = (1 - kCdL)^N$$

- Take (natural) logarithms:

$$\ln \frac{I}{I_0} = \ln(1 - kCdL)^N = N \ln(1 - kCdL)$$

A Very Handy Approximation

■ Consider the function $y = e^{-x}$

■ For $|x| \ll 1$:

$$e^{-x} \approx (1 - x)$$

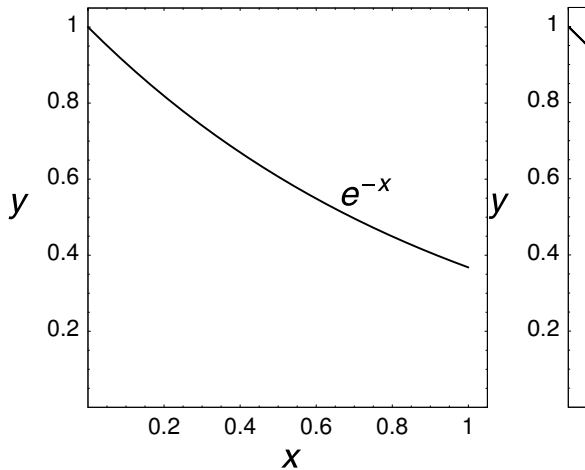
$$-x \approx \ln(1 - x)$$

■ This only works for e (≈ 2.71828).

For other numbers:

$$a^{-x} \approx (1 - x \ln a)$$

for $|x| \ll 1$



Many Thin Slices (contd.)

$$\ln \frac{I}{I_0} = N \ln(1 - kCdL)$$

- Our approximation: For $|x| \ll 1$:

$$(1 - x) \approx e^{-x}$$

$$\ln(1 - x) \approx -x$$

- Substituting:

$$\ln \frac{I}{I_0} = -NkCdL$$

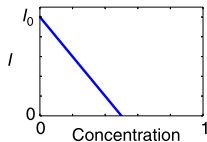
- $dL \cdot N = L$, total path length

$$\ln \frac{I}{I_0} = -C \cdot L \cdot k$$

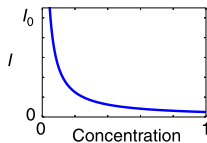
Clicker Question #3

How does transmitted light intensity change with concentration?

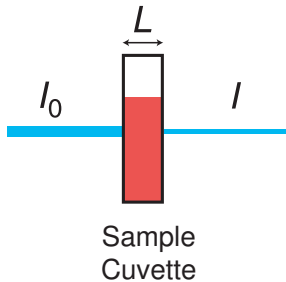
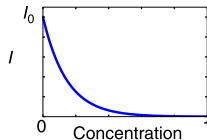
1 $I = I_0 - kC$



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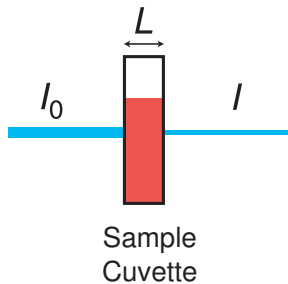
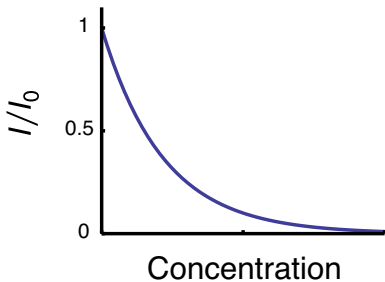


3 $I = I_0 e^{-kC}$



One correct answer!

The Beer-Lambert Law



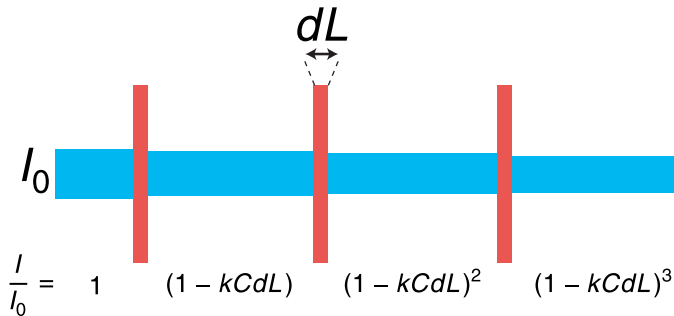
$$\ln \frac{I}{I_0} = -C \cdot L \cdot k$$

$$\log_{10}(x) = \ln(x) \log_{10}(e)$$

$$\log_{10} \frac{I}{I_0} = \ln \frac{I}{I_0} \log_{10}(e) = -C \cdot L \cdot k \log_{10}(e)$$

$$\log_{10} \frac{I_0}{I} = C \cdot L \cdot k \log_{10}(e) = C \cdot L \cdot \epsilon = A$$

A Quick Review of How We Got Here



■ For N slices: $\frac{I}{I_0} = (1 - kCdL)^N$

■ Take (natural) logarithms:

$$\ln \frac{I}{I_0} = \ln(1 - kCdL)^N = N \ln(1 - kCdL)$$

Units for the Extinction Coefficient

$$A = C \cdot L \cdot \epsilon$$

- A is dimensionless
- Most cuvettes have a path length of 1 cm, so it is convenient to use cm as the dimension of length.
- If concentration is expressed in molar units, then ϵ should have units of $M^{-1}cm^{-1}$, so that:
 $M \times cm \times M^{-1}cm^{-1}$ is dimensionless
- If concentration is expressed in units of mg/mL, then ϵ should have units of $cm^{-1}(mg/mL)^{-1} = cm^{-1}(mL/mg)$.
- If concentration is expressed as % (m/v) solute, then ϵ should have units of $cm^{-1}\%^{-1} = cm^{-1}(g/100mL)^{-1} = cm^{-1}(100mL/g)$.

Clicker Question #4

- Someone gives you a solution of a mystery compound and tells you that the extinction coefficient at 535 nm is $3 \text{ cm}^{-1}(\text{g/L})^{-1}$
- Using a 1 cm cuvette, the absorbance is 1.2.
- The concentration of the sample is:

1 0.04 mg/mL

2 0.4 mg/mL

3 4 mg/mL

4 0.04 g/mL

5 0.4 g/mL

6 4 g/mL

$$A = C \cdot L \cdot \epsilon$$

$$1.2 = C \times 1 \text{ cm} \times 3 \text{ cm}^{-1}(\text{g/L})^{-1}$$

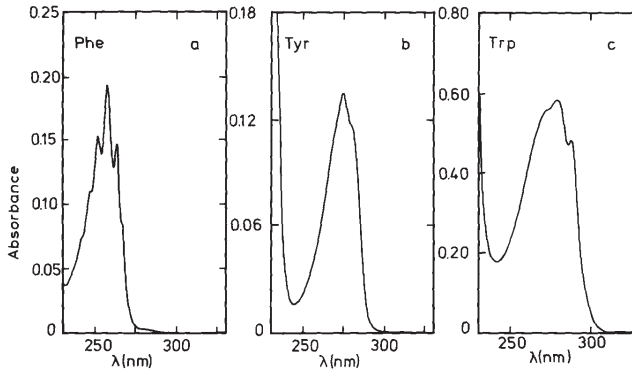
$$1.2 = C \times 3 (\text{g/L})^{-1}$$

$$C = 1.2 \div 3 (\text{g/L})^{-1}$$

$$C = 0.4 \text{ g/L} = 0.4 \text{ mg/mL}$$

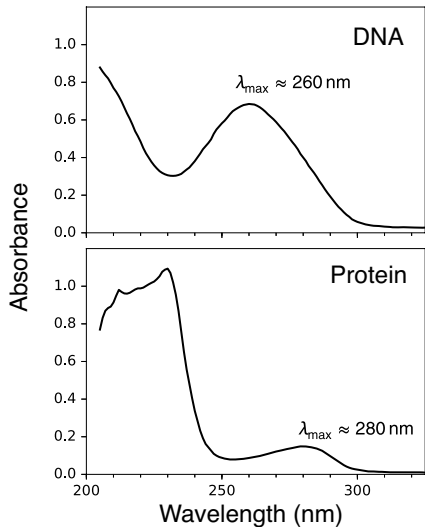
What if Solution Contains Multiple Compounds that Absorb Light?

- Peaks in UV-visible absorption spectra are quite broad:



- Peaks from different compounds often overlap.
- Absorption at a given wavelength may contain contributions from multiple compounds.

UV Absorption Spectra of Proteins and DNA

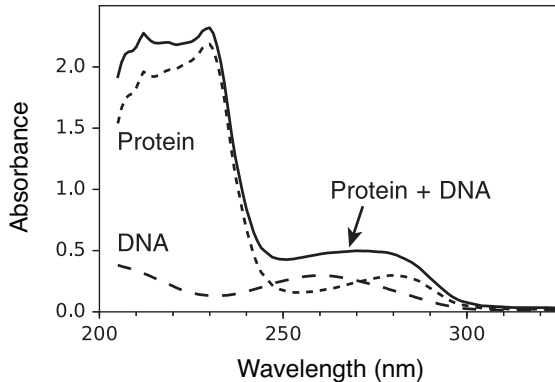


- DNA spectra do not depend much on sequence.
- Protein spectra do depend on amino acid composition, and a bit on three-dimensional structure.
- DNA and protein spectra, between 250 and 300 nm overlap extensively.
- Concentrations:
 - [DNA] \approx 0.03 mg/ml
 - [Protein] \approx 0.16 mg/ml

Spectra adapted from Spectrophotometry Handbook (2012) GE LifeScience

http://www.gelifesciences.com/file_source/GELS/Service%20and%20Support/Documents%20and%20Downloads/Handbooks/pdfs/Spectrophotometry.pdf

Spectra of DNA, Protein and a Mixture



- Absorbances of different components add.
- Assumes components don't interact.
- Can we interpret absorbance of mixtures?

Estimating Concentrations of Protein and DNA in a Mixture

- Between 250 and 300 nm

For Protein: $\lambda_{\max} \approx 280 \text{ nm}$

For DNA: $\lambda_{\max} \approx 260 \text{ nm}$

- At 260 nm (assuming 1-cm cuvette):

$$A_{260} = [\text{Protein}] \cdot \epsilon_{260}^{\text{Protein}} + [\text{NA}] \cdot \epsilon_{260}^{\text{NA}}$$

- At 280 nm:

$$A_{280} = [\text{Protein}] \cdot \epsilon_{280}^{\text{Protein}} + [\text{NA}] \cdot \epsilon_{280}^{\text{NA}}$$

- If all four extinction coefficients are known, and we measure A_{260} and A_{280} , we have two equations in two unknowns.

Solve for [Protein] and [NA].

- What could go wrong?

Outline of Experiment

- Two samples:
 - A pure protein: Bovine serum albumin (BSA)
 - An *E. coli* extract, containing lots of proteins and nucleic acids
- Direct UV absorption measurements at 260 and 280 nm
 - For BSA, estimate [Protein] from A_{280} and known extinction coefficient.
 - For both samples, estimate [Protein] and [NA] from extinction coefficients for “typical” proteins and nucleic acids.
- Bradford dye-binding assay
 - Use BSA to establish a standard curve, using [BSA] determined from A_{280}
 - Independent estimate [Protein] in *E. coli* extract, to be compared with estimate from $A_{280} : A_{260}$